

# A study of essential oil extraction and antioxidant activity of patchouli (*Pogostemon cablin*) using supercritical carbon dioxide

Soon Hong Soh<sup>1,2</sup>, Shuchi Agarwal<sup>1</sup>, Sundaramurthy Jayaraman<sup>1</sup>, Ming Tan Tham<sup>2</sup>,  
Cindy Lai Yeng Lee<sup>2\*</sup>, and Akshay Jain<sup>1\*</sup>

<sup>1</sup> EWT Centre of Innovation, Ngee Ann Polytechnic, 535 Clementi Road Singapore 599489

<sup>2</sup> Newcastle University, Singapore, 537 Clementi Road, Singapore 599493

## Abstract

This study investigates the extraction of patchouli oil using supercritical carbon dioxide (SC-CO<sub>2</sub>). The patchouli plants utilized in this study were obtained from Indonesia. The effect of pressure on yield, antioxidant activity and total phenolic content of the extracts was investigated at operating pressures ranging from 100 – 200 bar. The extract yield, antioxidant activity (evaluated by 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay) and total phenolic content were compared against conventional steam distillation. SC-CO<sub>2</sub> extraction obtained extract yields and total phenols in the range of 3.41 to 3.95% and 1.869 to 2.756 mg GAE/g respectively, which were higher than steam distillation (1.13% and 1.677 mg GAE/g). Furthermore, the SC-CO<sub>2</sub> extracts exhibited IC<sub>50</sub> (effective concentration to reduce 50% of DPPH free radicals) values in the range of 0.581 mg/ml to 1.538 mg/ml, which were better than the IC<sub>50</sub> value exhibited by steam distilled patchouli oil (> 4 mg/ml).

## Introduction

*Pogostemon cablin* (patchouli) is an aromatic herb that is extensively cultivated in Brazil, China and Indonesia for its essential oil. The essential oil of patchouli plant is known to have great business potential in the international market due to its unique flavour, fragrance and biological activities [1]. In the perfumes and cosmetics industry, patchouli oil is a vital ingredient in exotic perfumes offering a rich and spicy fragrance. Its fixative properties provides a strong lasting character to perfumes when blended with other essential oils. It is also used as a perfume in its own right [2]. Furthermore, the fixative properties of patchouli oil assists in limiting evaporation and promoting firmness, allowing it to be valuable in the manufacturing of soaps, scents, body lotions and detergents. In aromatherapy, it is used to soothe nerves, control appetite, and relieve depression and stress. Moreover, patchouli oil is on the FDA's (Food and Drug Administration) list of substances approved for human consumption as a natural additive for food flavouring [3]. In food industries, patchouli oil is extensively used as a flavour ingredient in major food products including alcoholic and non-alcoholic beverages [4]. Very low concentration (2 mg/kg) of this oil is used to flavour foods, beverages, candy and baked products [5].

Indonesia is the largest producer of patchouli oil, accounting for over 80 percent of its total annual production [6]. Patchouli oil extraction in Indonesia is generally carried out using conventional methods which limits their production capacity [7]. Conventional steam distillation of dried patchouli leaves have drawbacks such as long extraction time, low yield and adverse effects of high temperature [7,8]. High temperature is detrimental to heat sensitive compounds present in essential oil as it may cause a chemical alteration, resulting in a different

flavour and fragrance profile. For instance, fatty aliphatic aldehydes, terpenic hydrocarbons, as well as terpenoid aldehydes, in citrus oils are thermally unstable and are readily oxidized by atmospheric oxygen which gives rise to undesirable formation of malodorous carboxylic acids [9]. In this regard, the application of supercritical fluids is of considerable interest. SC-CO<sub>2</sub> extraction involves mild operating temperature, while being organic solvent free, is especially beneficial in cosmetics, foods, and pharmaceutical products where there are more stringent requirements.

Limited studies were conducted on SC-CO<sub>2</sub> extraction of patchouli oil. For instance, Donelian et al. [10] studied the yield and chemical composition of essential oil obtained by SC-CO<sub>2</sub> extraction from patchouli leaves cultivated in Brazil. Liu et al. [11] studied the yield of concrete and oil obtained by SC-CO<sub>2</sub> extraction from patchouli leaves and stems from China. In this work, the objective is to study SC-CO<sub>2</sub> extraction of essential oil from patchouli plant (Indonesia), to analyse the effect of pressure on extract yield, antioxidant activity and total phenolic content and to compare extraction results with conventional steam distillation.

## **Materials and methods**

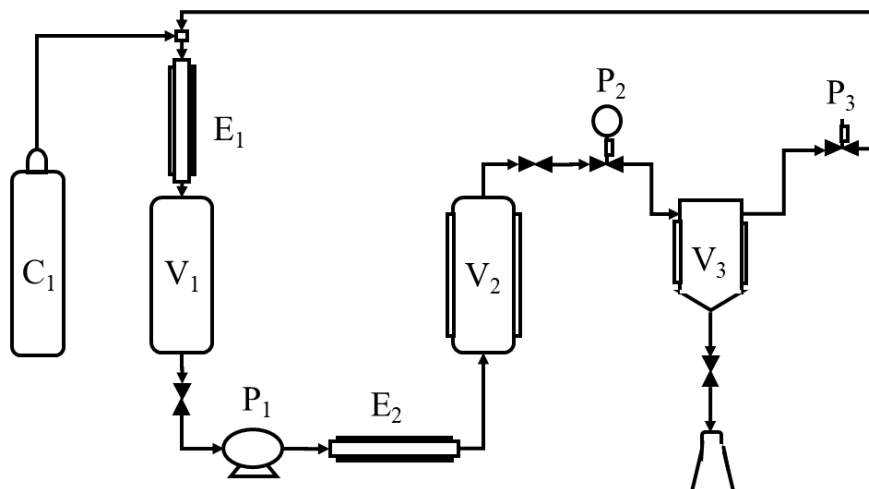
### *Materials*

Dried patchouli (*Pogostemon cablin*) plant with a moisture content of  $10.60 \pm 0.07\%$ , measured using a halogen heating moisture analyser (MOC63u UniBloc, Shimadzu), was obtained from Lam Seng Hang Co. Pte Ltd (Indonesia). The leaves and stems were separated initially followed by grinding in an electric blender. The ground material was then passed through a 0.6 mm stainless steel sieve. Samples were packed and stored at room temperature until utilisation. SC-CO<sub>2</sub> extraction experiments were carried out with a mixture comprising of 1:1 ratio of ground leaves and stems. Liquid CO<sub>2</sub> with a purity of 99.5% was obtained from Air Liquide Singapore Pte Ltd. Ethanol 96% (technical grade) was obtained from Quality Reagent Chemical. Methanol (analytical grade) was obtained from Fischer Scientific Pte Ltd (Singapore). Anhydrous sodium carbonate, sodium sulphate and 2,2-diphenyl-1-picrylhydrazyl (DPPH) were procured from Sigma-Aldrich Pte Ltd (Singapore). Folin Ciocalteu's phenol reagent and gallic acid were procured from Merck Pte Ltd (Singapore) and used as obtained.

### *Supercritical CO<sub>2</sub> extraction*

SC-CO<sub>2</sub> extractions were carried out using a customized supercritical fluid extractor with CO<sub>2</sub> recycle system (Model SFE 1000 System, Waters Corporation, USA). Figure 1 shows a schematic diagram of the process. Ground samples were loaded and packed with glass beads. The loaded sample was heated until the desired temperature was reached. Liquid CO<sub>2</sub> was pressurized by a cooled liquid pump to the desired extraction pressure, followed by a preheating stage where it was heated to the extraction temperature. The pressure of the extraction vessel was controlled by an automated backpressure regulator (ABPR). The pressure of the collection vessel was maintained below critical pressure by a manual backpressure regulator (MBPR), for the recycling of CO<sub>2</sub>. At this stage, the CO<sub>2</sub> reverts to its gaseous state and the plant extracts precipitate at the bottom of the vessel. The temperature in the extraction and collection vessels were maintained using band heaters. The plant extracts were collected from the collection vessel at the end of each run. After collection, the pipeline of the system was purged with CO<sub>2</sub> to recover residues. Small amounts of extracts that remained within the walls of the collection vessel were rinsed down with ethanol before evaporating using a rotary evaporator (Heidolph, Germany). The extracts were weighed and stored in a refrigerator for further analysis. The extract yield obtained from each experiment were calculated using the following equation:

$$\text{Extract yield (\%)} = \frac{\text{Mass of extract (g)}}{\text{Mass of sample (g)}} \times 100\% \quad (1)$$



**Figure 1:** Supercritical fluid extraction setup C<sub>1</sub>: Compressed CO<sub>2</sub> cylinder; E<sub>1</sub>: Condenser; E<sub>2</sub>: Electric preheater; P<sub>1</sub>: High pressure liquid pump; P<sub>2</sub>: Automated backpressure regulator; P<sub>3</sub>: Manual backpressure regulator V<sub>1</sub>: CO<sub>2</sub> recycler; V<sub>2</sub>: Extraction vessel; V<sub>3</sub>: Collection vessel

#### Steam distillation

100 g of dried patchouli leaves were placed in a biomass flask and subjected to steam distillation for 5 h. Patchouli oil was separated from water using a separatory funnel. Residual water content was removed from patchouli oil by drying with anhydrous sodium sulphate. The dehydrated patchouli essential oil was weighed and stored in a refrigerator for further analysis.

#### Antioxidant activity analysis

Antioxidant activity was analysed by free radical scavenging activity of the patchouli extract towards 2, 2-diphenyl-1-picrylhydrazyl (DPPH) [12,13]. A range of concentration from 0.25 to 4 mg/ml of the plant extracts were prepared with methanol. Plant extract solutions (2 ml each) of the different concentrations were mixed with 3 ml of freshly prepared 0.004% (w/v) DPPH methanol solution and shaken well before incubating in dark and ambient conditions for 30 minutes. The absorbance of the sample was then analysed and recorded at 517 nm using an ultraviolet and visible (UV-Vis) spectrophotometer (UV-1800, Shimadzu). The percentage of radical scavenging activity was calculated using the following equation:

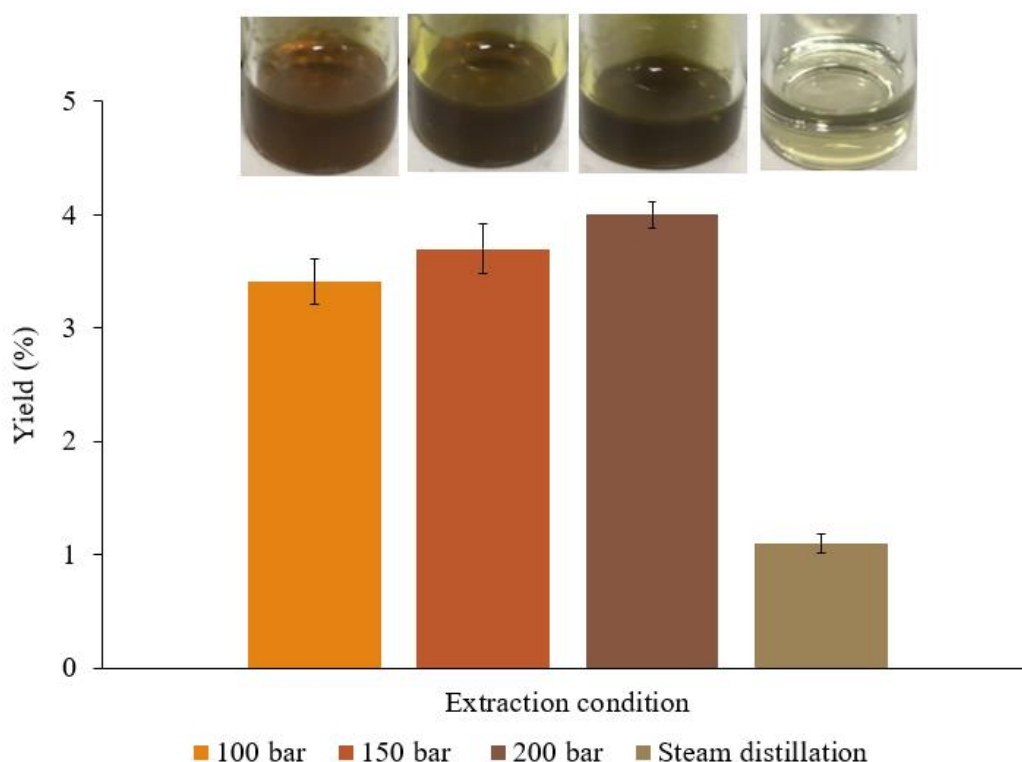
$$I\% = \frac{A_b - A_s}{A_b} \times 100 \quad (2)$$

Where  $A_b$  and  $A_s$  are the absorbance values of the DPPH blank solution and tested samples respectively. Lower absorbance values of sample indicated higher DPPH radical scavenging activity. IC<sub>50</sub> value (mg/ml) is the effective concentration of essential oil at which DPPH radicals were scavenged by 50%. This was estimated by interpolation and linear regression.

#### Total phenolic content analysis

Total phenolic content was analysed using the Folin Ciocalteu method with some modifications [14]. 200 µl of properly diluted extract (10% w/v in methanol) or a standard solution of varying concentrations was mixed with 400 µl of Folin Ciocalteu's phenol reagent. The solution was diluted to a total volume of 4.6 ml and thoroughly mixed. After incubating for 10 min at room temperature, 1 ml of 20% sodium carbonate (Na<sub>2</sub>CO<sub>3</sub>) solution was added to the solution and then instantly mixed, centrifuged and incubated for 2 h. The absorbance was measured at 760

nm using an UV-Vis spectrophotometer. Gallic acid was used as the standard and the total phenolic content of samples were expressed in milligram gallic acid equivalent per gram of extract (mg GAE/g).

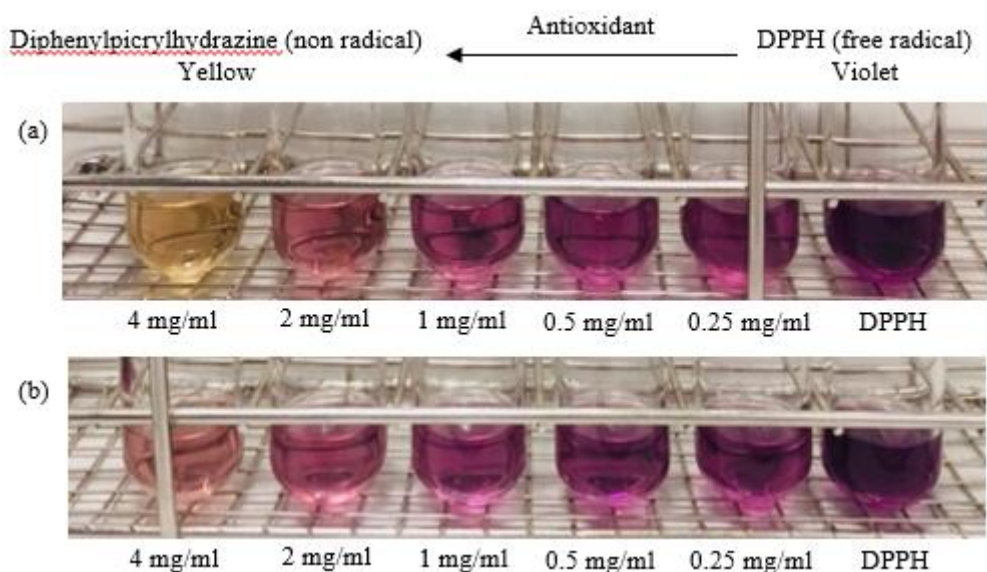


**Figure 2:** Extract profile and yield under different pressures of SC-CO<sub>2</sub> extraction and by steam distillation

## Results and discussion

### *Extract yield*

SC-CO<sub>2</sub> extractions were conducted at pressures ranging from 100 – 200 bar. The extractions were fixed at 40°C. Figure 2 shows the extract yield under different conditions of SC-CO<sub>2</sub> extraction and comparison with extraction by steam distillation. An increase in pressure resulted in higher yield, which was attributed to an increase in SC-CO<sub>2</sub> density and solvation power at higher pressure. Moreover, in this study, the extract yield by SC-CO<sub>2</sub> extraction is clearly higher than that obtained by steam distillation which is consistent with results reported in similar work [10,12,15]. Steam distillation lacks the capability to recover non-volatile compounds. For instance, sclareol, an important component of clary sage, is usually recovered in only very small quantities in steam distillation due to its very high boiling point [9]. On the other hand, the selective nature of SC-CO<sub>2</sub> extraction allows it to extract a wide range of compounds by fine-tuning operating conditions and altering the properties of SC-CO<sub>2</sub>. As the density of SC-CO<sub>2</sub> increased, more non-volatiles were extracted. Therefore, the ability to extract non-volatile compounds enables SC-CO<sub>2</sub> extraction to achieve a higher extract yield than steam distillation. However, this also meant that undesirable non-volatile compounds such as cuticular waxes were co-extracted in SC-CO<sub>2</sub> extraction [16]. In addition to extract yield, Figure 2 shows the profiles of the extracted patchouli oils from steam distillation and SC-CO<sub>2</sub> extraction at different pressures. Despite having higher yields, the dark brown colour extracts observed from SC-CO<sub>2</sub> extraction indicated some co-extraction of heavier, non-volatile compounds as compared to the pale yellow volatile oil obtained from steam distillation [17]. Furthermore, the colour of the extract became more intense as SC-CO<sub>2</sub> extraction pressure increased due to the presence of more non-volatile compounds [18].



**Figure 3:** Extract solution at different concentrations (0.25 – 4 mg/ml) mixed with DPPH (SC-CO<sub>2</sub> extraction pressure: (a) 200 bar and (b) 150 bar)

#### *Antioxidant activity and total phenolic content*

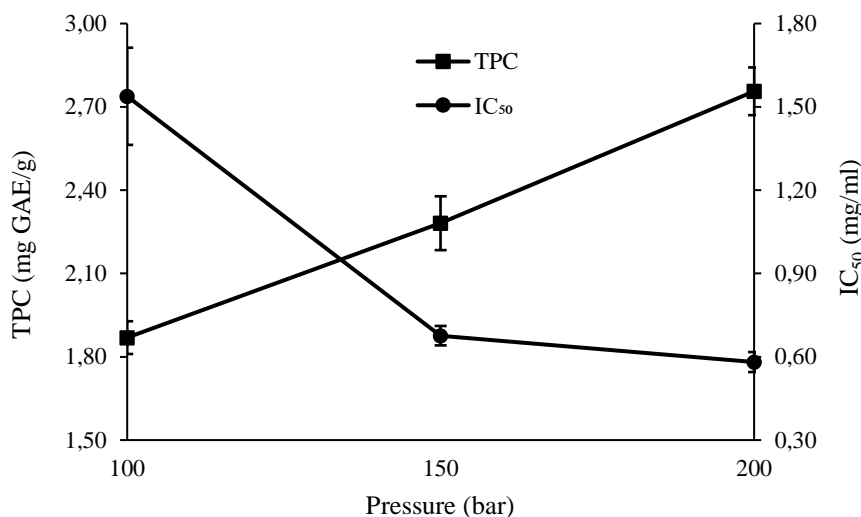
Antioxidant activity of extracts were evaluated by a DPPH radical-scavenging assay that predicted the ability to quench free radicals. The radical scavenging activity was determined by measuring the degree of absorbance quenching for varying sample concentrations. Figures 3(a) and (b) show the degree of absorbance quenching at different sample concentrations obtained from SC-CO<sub>2</sub> extraction at 200 bar and 150 bar respectively.

Table 1 displays the IC<sub>50</sub> values and total phenols under different pressures of SC-CO<sub>2</sub> extraction and by steam distillation. Experimental results showed IC<sub>50</sub> values ranging from 0.581 – 1.538 mg/ml and total phenols ranging from 1.869 – 2.756 mg GAE/g under different pressures of SC-CO<sub>2</sub> extraction. Lower IC<sub>50</sub> values indicated better antioxidant activity. Table 1 shows that SC-CO<sub>2</sub> extraction resulted in better antioxidant activity and more total phenols than steam distillation (> 4 mg/ml and 1.677 mg GAE/g). This highlights the capability of SC-CO<sub>2</sub> extraction of recovering non-volatile, polar bioactive compounds as opposed to steam distillation, which is only able to extract volatile oils. Therefore, the higher yields attained in SC-CO<sub>2</sub> extraction can be further explained by the co-extraction of bioactive compounds on top of the volatile compounds generally obtained by steam distillation. Steam distilled patchouli oil have lower antioxidant activity due to low content of active components. During steam distillation of patchouli oil, the antioxidant activity is partially lost. Steam distilled oils contain mostly volatile compounds, which have generally low antioxidant activity and can be limitedly applied in food industry as authentic antioxidants [19]. The characteristics of the target antioxidant compounds must be considered in the selection of extraction method. The active components extracted from plants depend on heat, light, oxygen, and a number of other factors. The prolonged exposure of the patchouli oil to heat and light in steam distillation had degraded these active compounds and caused major deterioration to the antioxidant activity [12,19,20]. SC-CO<sub>2</sub> extraction was performed in a closed extraction vessel without the presence of light and oxygen, therefore this minimized the degradation reactions of active

compounds. Furthermore, SC-CO<sub>2</sub> extraction was performed at mild temperatures as compared to steam distillation and therefore degradation of heat sensitive compounds was reduced.

**Table 1:** IC<sub>50</sub> values and total phenols obtained from extracts under different pressures of SC-CO<sub>2</sub> extraction and by steam distillation

Extraction	IC <sub>50</sub> (mg/ml)	Total phenols (mg GAE/g)
100 bar	1.538 ± 0.175	1.869 ± 0.059
150 bar	0.676 ± 0.035	2.281 ± 0.097
200 bar	0.581 ± 0.036	2.756 ± 0.086
Steam distillation	> 4	1.677 ± 0.068



**Figure 4:** Effect of pressure on antioxidant activity and total phenols obtained by SC-CO<sub>2</sub> extraction (TPC = total phenolic content)

Figure 4 shows the main effect plot of pressure on the antioxidant activity and total phenols obtained by SC-CO<sub>2</sub> extraction. It was observed that the total phenols increased at higher pressures. Similar to extract yield, the observation was due to the improved density of SC-CO<sub>2</sub> at higher pressures, which improved the solvation power of the solvent. As the solvent density increased at higher pressures, the distance between the molecules decreased. Therefore, interaction between solutes and CO<sub>2</sub> increased, permitting higher specific extraction of polar active compounds [12]. In addition, Figure 4 shows that a rise in pressure displayed positive effect on the antioxidant activity of the extract. This is reflected by the enriched extraction of polyphenols, amongst other bioactive compounds that may be present, at higher pressures as these compounds contribute to antioxidant activity [13,21].

## Conclusion

SC-CO<sub>2</sub> extraction of essential oil from Indonesian based patchouli plant was studied and compared against conventional steam distillation. The effect of pressure on extract yield, antioxidant activity and total phenolic content was investigated. SC-CO<sub>2</sub> extraction offers a selective extraction which can be targeted towards recovering compounds of interest by changing operating parameters. The extract yield, antioxidant activity and total phenols were all improved at higher pressures. SC-CO<sub>2</sub> extraction displayed higher yields of extract and phenols, with better antioxidant activities as compared to steam distillation. Due to its solvent free and low temperature processing nature, enhanced yield and bioactive compounds with therapeutic benefits, SC-CO<sub>2</sub> extraction can be a better alternative to conventional methods for

application. Further studies on the optimization of the process parameters and techno-economic evaluation of the SC-CO<sub>2</sub> extraction process for patchouli oil will be carried out for large-scale production.

### Acknowledgements

This work was supported by Ministry of Education (Singapore) Funding MOE2015-TIF-2-G-051 and Newcastle University PhD scholarship for Singapore. The authors would like to express their gratitude to Lam Seng Hang Co. Pte Ltd for arranging the timely collection of the samples used in this work.

### References

- [1] SWAMY M.K., SINNIAH U.R., A comprehensive review on the phytochemical constituents and pharmacological activities of *Pogostemon cablin* Benth.: An aromatic medicinal plant of industrial importance, *Molecules*. 20, 2015, p. 8521–8547
- [2] RAMYA H.G., PALANIMUTHU V., RACHNA S., An introduction to patchouli (*Pogostemon cablin* Benth.) - A medicinal and aromatic plant: It's importance to mankind, *Agric. Eng. Int. CIGR J.* 15, 2013, p. 243–250
- [3] FEDERAL REGULATIONS CODE. Food and Drugs Administration, from the U.S. Government Printing Office via GPO Access [CITE: 21CFR172.510]. U.S.A., 3, 2002, p. 49–52
- [4] DAS K., Patchouli (*Pogostemon cablin* Benth) oils, in: *Essent. Oils Food Preserv. Flavor Saf.*, 2015, p. 633–639
- [5] BAUER K., GARBE D., SURBURY H., *Common Fragrance and Flavour Materials: Preparation, Properties and Used*, third ed., Wiley-VCH, Weinheim, 1997, p. 205
- [6] TUCKER A.O., DEBAGGIO T., *The Encyclopedia of Herbs: A comprehensive reference to herbs of flavour and fragrance*, second ed., Timber Press, Portland, 2009, p. 404
- [7] KUSUMA H.S., MAHFUD M., The extraction of essential oils from patchouli leaves (*Pogostemon cablin* benth) using a microwave air-hydrodistillation method as a new green technique, *RSC Adv.* 7, 2017, p. 1336–1347
- [8] YAHYA A., YUNUS R.M., Influence of sample preparation and extraction time on chemical composition of steam distillation derived patchouli oil, *Procedia Eng.* 53, 2013, p. 1–6
- [9] BASER K., BUCHBAUER G., *Handbook of Essential oils: Science, Technology, and Applications*, CRC Press, Boca Raton, 2010, p. 96
- [10] DONELIAN A., CARLSON L.H.C., LOPES T.J., MACHADO R.A.F., Comparison of extraction of patchouli (*Pogostemon cablin*) essential oil with supercritical CO<sub>2</sub> and by steam distillation, *J. Supercrit. Fluids.* 48, 2009, p. 15–20
- [11] LIU Y., SUN B., SHI H., ZHENG F., Extraction of concrete of patchouli stem and leaf with supercritical carbon dioxide, *Huagong Xuebao/Journal Chem. Ind. Eng.* 59, 2008, p. 791–795.
- [12] SODEIFIAN G., SAJADIAN S.A., Investigation of essential oil extraction and antioxidant activity of *Echinophora platyloba* DC. using supercritical carbon dioxide, *J. Supercrit. Fluids.* 121, 2017, p. 52–62
- [13] MOHAMMED, N.K., MANAP, M.Y.A., TAN, C.P., MUHIALDIN, B.J., ALHELLI, A.M., HUSSIN, A.S.M., The Effects of Different Extraction Methods on Antioxidant Properties, Chemical Composition, and Thermal Behavior of Black Seed (*Nigella sativa* L.) Oil. *Evid Based Complement Alternat Med.*, 2016, Article ID 6273817
- [14] SINGLETON V.L., ROSSI J.A., Colorimetry of Total Phenolics with Phosphomolybdic-Phosphotungstic Acid Reagents, *Am. J. Enol. Vitic.* 16, 1965, p. 144–158
- [15] HERZI N., BOUAJILA J., CAMY S., CAZAUX S., ROMDHANE M., CONDORET J.S., Comparison between Supercritical CO<sub>2</sub> Extraction and Hydrodistillation for Two

Species of Eucalyptus: Yield, Chemical Composition, and Antioxidant Activity, J. Food Sci. 78, 2013, p. C667-72

[16] GASPAR F., Extraction of Essential Oils and Cuticular Waxes with Compressed CO<sub>2</sub>: Effect of Extraction Pressure and Temperature, Ind. Eng. Chem. Res. 41, 2002, p. 2497–2503

[17] SAHA S., WALIA S., KUNDU A., SHARMA K., SINGH J., TRIPATHI B., RAINA A., Compositional and functional difference in cumin ( *Cuminum cyminum* ) essential oil extracted by hydrodistillation and SCFE, Cogent Food Agric. 2, 2016, Article ID 1143166

[18] GOMES P.B., MATA V.G., RODRIGUES A.E., Production of rose geranium oil using supercritical fluid extraction, J. Supercrit. Fluids. 41, 2007, p. 50–60

[19] INANC T., MASKAN M., Testing the antioxidant effect of essential oils and BHT on corn oil at frying temperatures: A response surface methodology, JAOCS, J. Am. Oil Chem. Soc. 90, 2013, p. 1845–1850

[20] WONG Y.M., SIOW L.F., Effects of heat, pH, antioxidant, agitation and light on betacyanin stability using red-fleshed dragon fruit (*Hylocereus polyrhizus*) juice and concentrate as models, J. Food Sci. Technol. 52, 2015, p. 3086–3092

[21] VLADIMIR-KNEZEVI S., BLAZEKOVI B., Plant Polyphenols as Antioxidants Influencing the Human Health, Phytochem. as Nutraceuticals – Glob. Approaches to Their Role Nutr. Heal., 2012, p. 155–180